

# High rhamnolipids production correlates with a high intracellular *R*- to *S*-specific enoyl-CoA hydratase activity

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## Abstract

Rhamnolipids (RLs) are multipurpose surface-active molecules produced from rhamnose and *R*-3-hydroxyalkanoate ( $C_{10\pm 2}$ ) precursors by the pathogenic bacterium *Pseudomonas aeruginosa*.

Interestingly, 3-hydroxyalkanoate precursors of RLs are exclusively in the *R*-form and we recently reported that they originate from enoyl-CoA ( $C_{10\pm 2}$ ) intermediates of  $\beta$ -oxidation diverted by the action of *R*-specific enoyl-CoA hydratase (*R*-ECH), called RhIYZ.

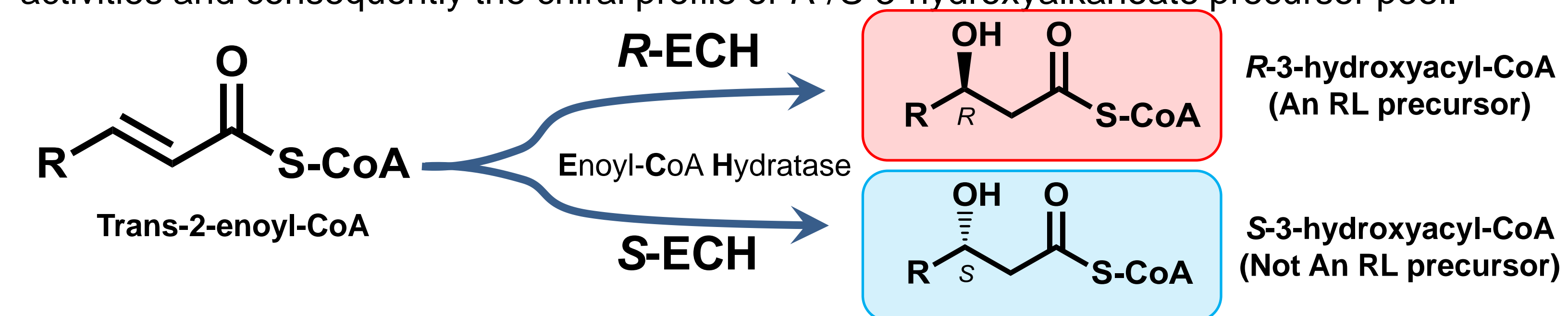
*S*- and *R*-ECH catalyze the stereospecific hydration of 2-enoyl-CoA into the corresponding *S*- and *R*-3-hydroxyacyl-CoA, respectively.

In this study, we investigated the correlation between the net *R*/*S*-ECH activity of *P. aeruginosa* PA14 in comparison with different bacteria under various culture conditions using our recently developed chiral HPLC method coupled with tandem mass spectrometry (MS) for the stereospecific and quantitative analysis of *S*/*R*-3-hydroxyalkanoates.

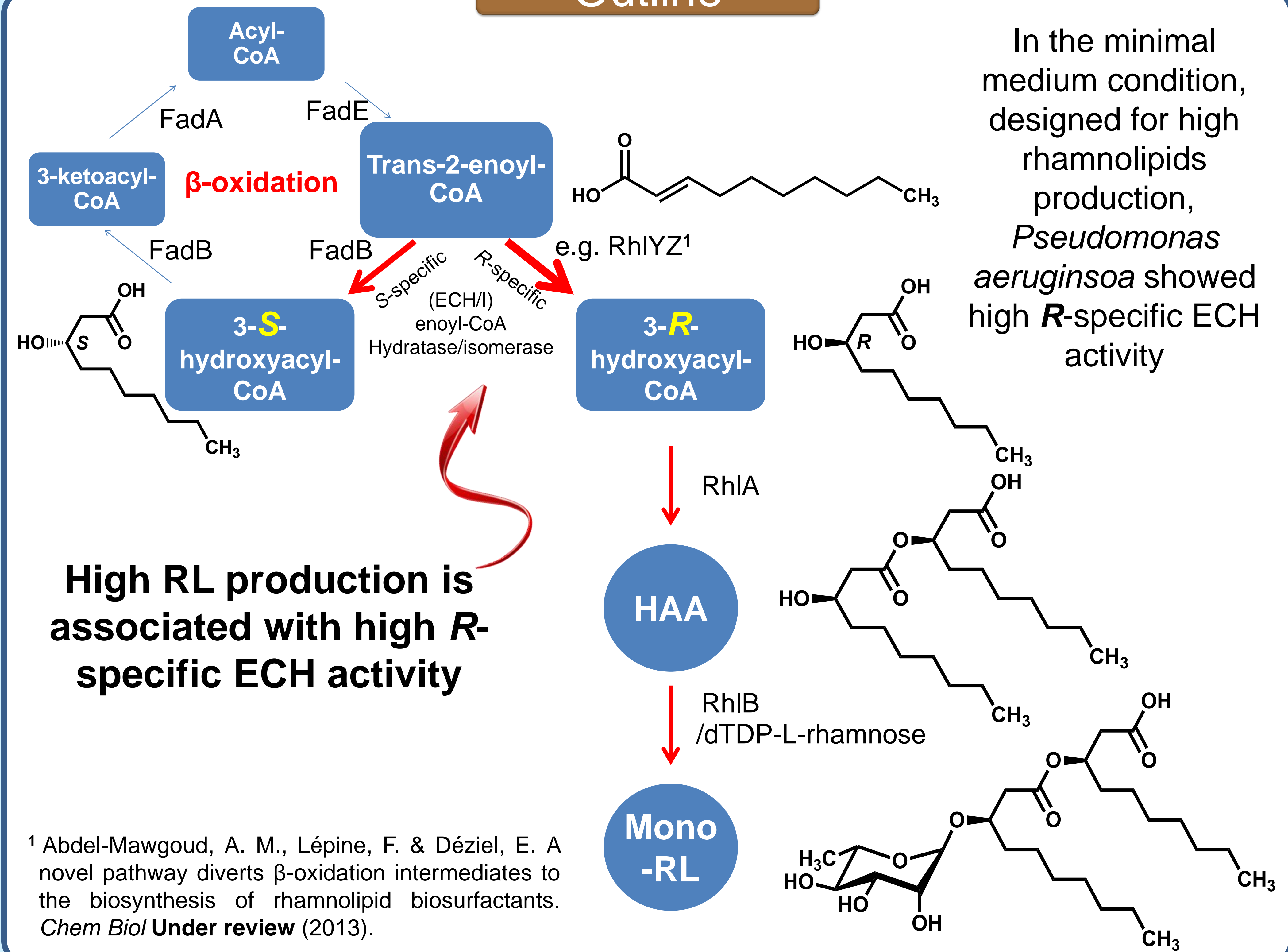
We show that, in mineral salts medium designed for high RLs production, the net ECH activity of *P. aeruginosa* is almost exclusively of the *R*-specific type (*R*-ECH). On the other hand, in rich medium conditions characterized by low RLs production, a nearly equal *S*- to *R*-ECH activities was observed.

We also evaluated the net ECH activity in two other bacteria that are candidates for the heterologous expression of rhamnolipids genes, namely, *Escherichia coli* and *Pseudomonas putida*, and found that the former is of mainly *S*-ECH type whether in rich or minimal media, while the latter, like *P. aeruginosa*, is of equal *R*/*S*-ECH in rich medium and of *R*-ECH in minimal medium.

These observations could explain the reported failures to express RLs production in *E. coli* and its success in *P. putida*. This study reveals an important analytical tool for evaluation of the candidacy of different hosts for the heterologous expression of RLs, preferred in non-pathogenic hosts for commercial purposes, based on the nature of the net *R*/*S*-ECH activities and consequently the chiral profile of *R*/*S*-3-hydroxyalkanoate precursor pool.



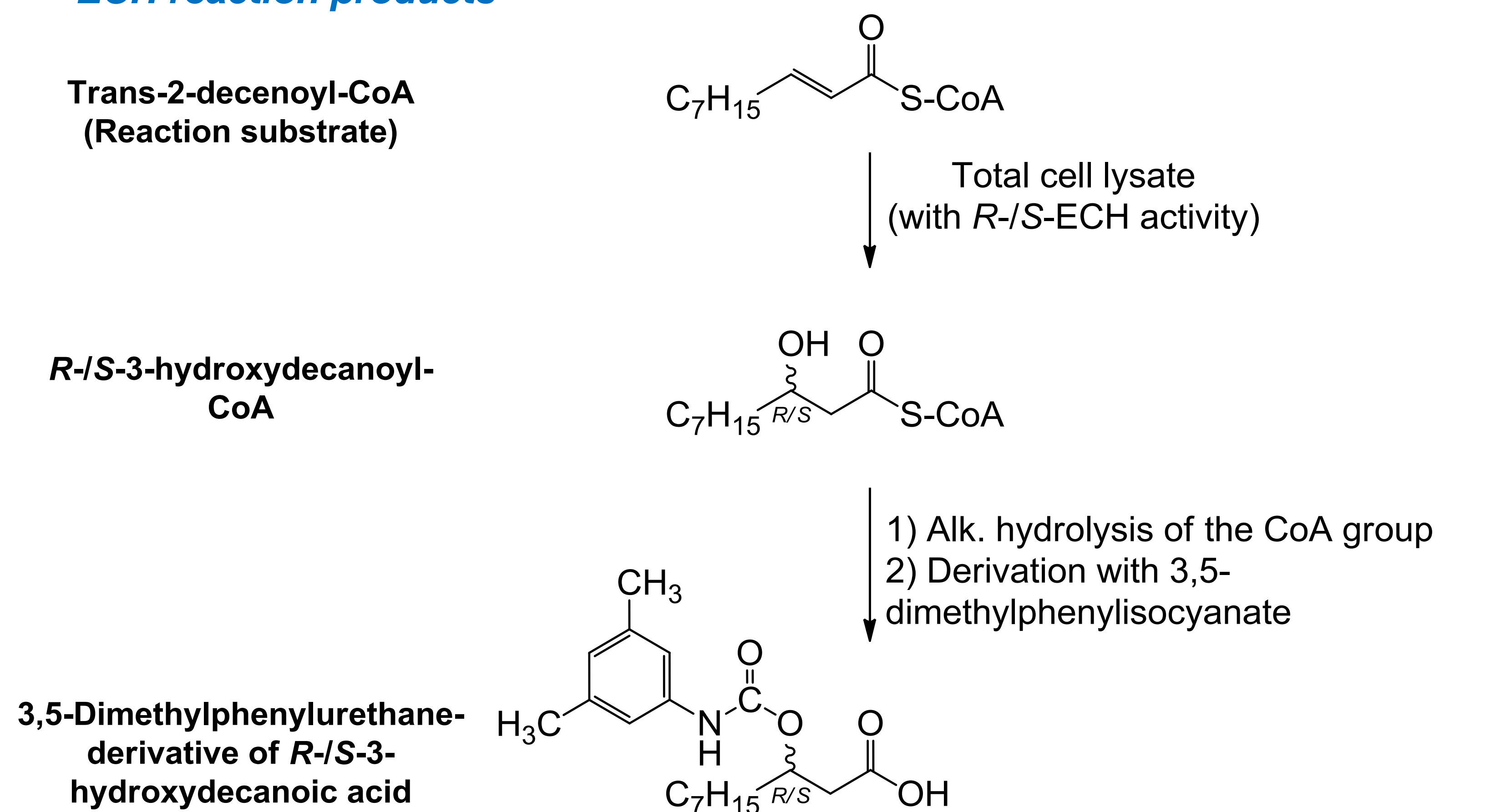
## Outline



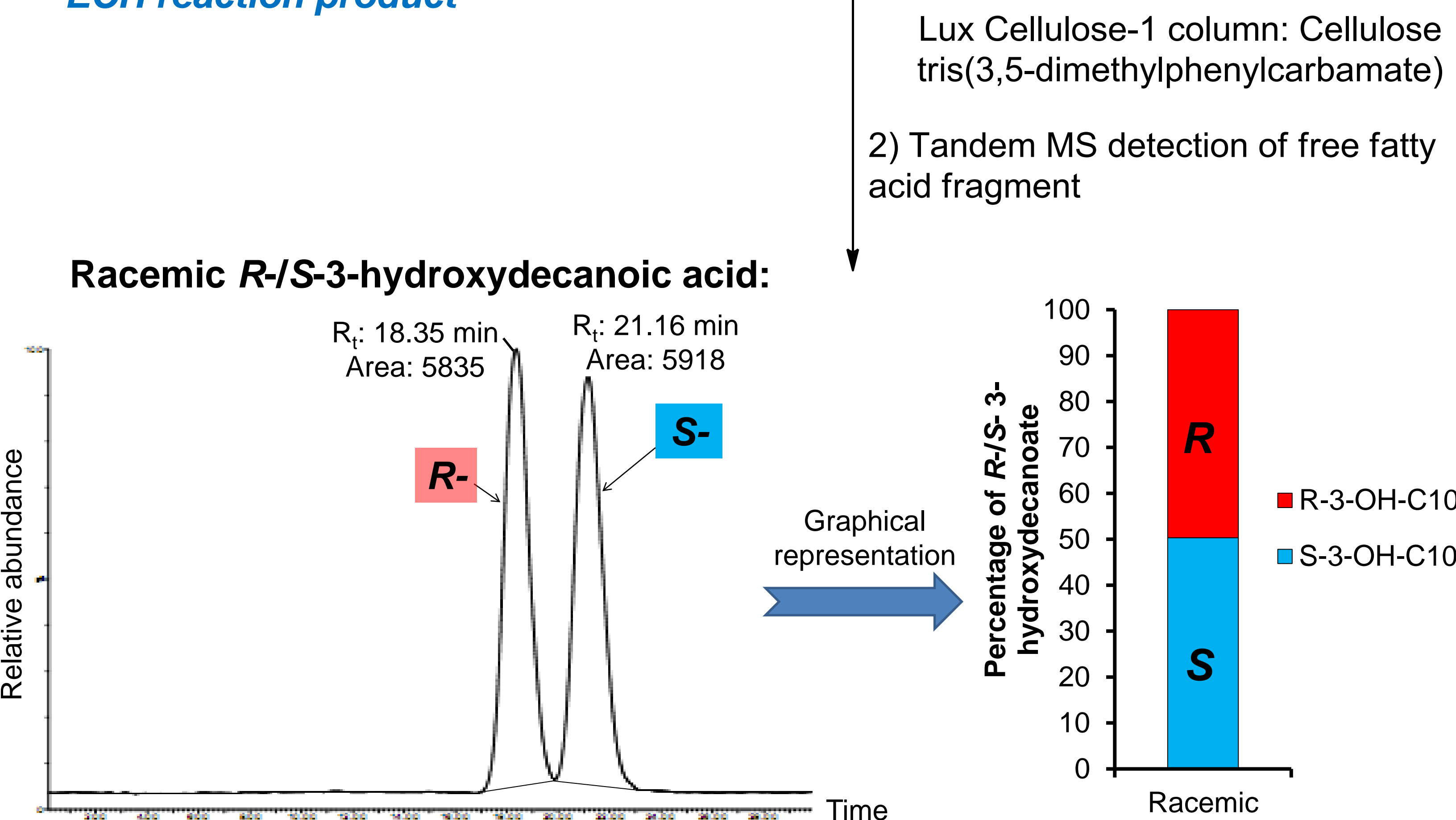
## Methodology

### HPLC/MS-MS method for estimation of *R*/*S*-ECH activities <sup>2</sup>

#### *In vitro* ECH reaction using clarified total cell lysate and derivatization of ECH reaction products



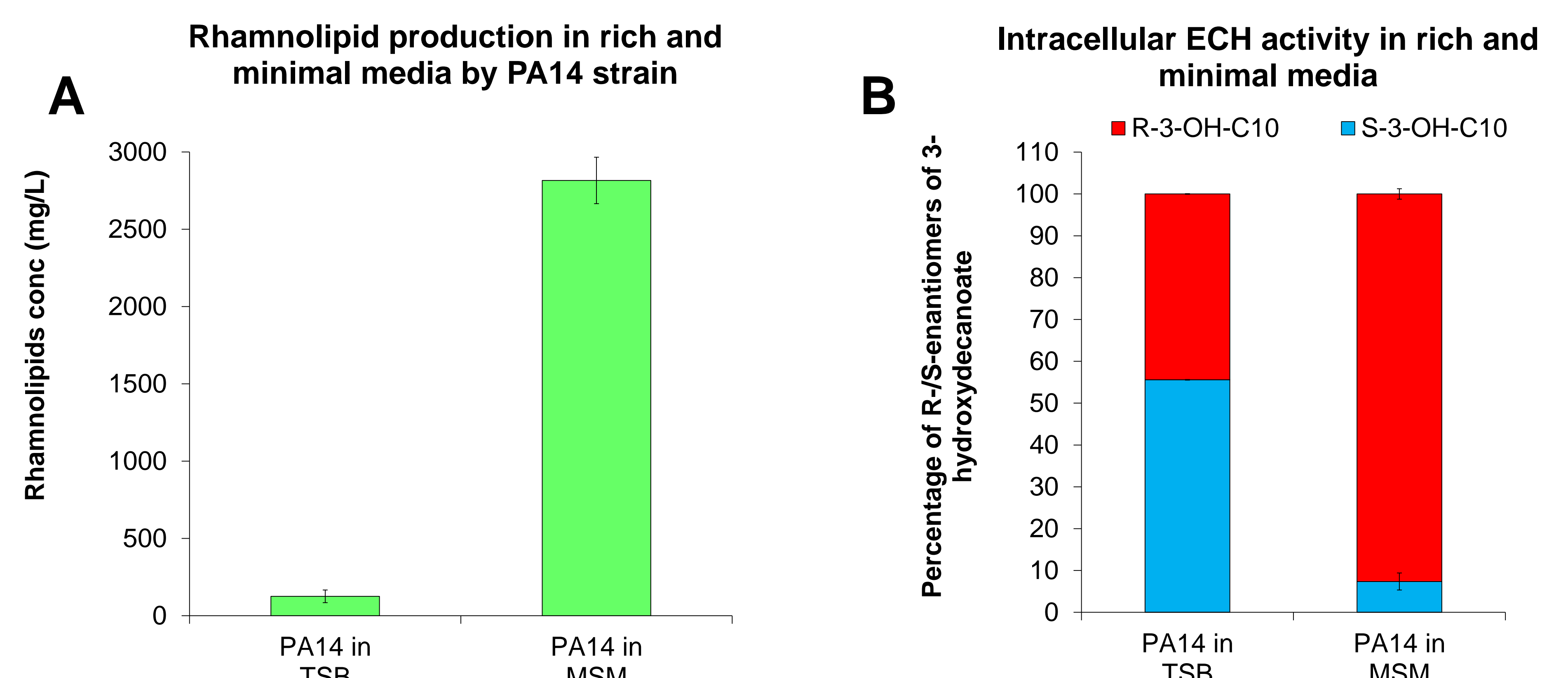
#### Chiral chromatographic separation and tandem MS detection of derivatized ECH reaction product



<sup>2</sup> Abdel-Mawgoud, A. M., Lépine, F. & Déziel, E. A chiral high-performance liquid chromatography-tandem mass spectrometry method for the stereospecific analysis of enoyl-coenzyme A hydratases/isomerases. *Journal of Chromatography A* 1306, 37-43 (2013).

## Results-1

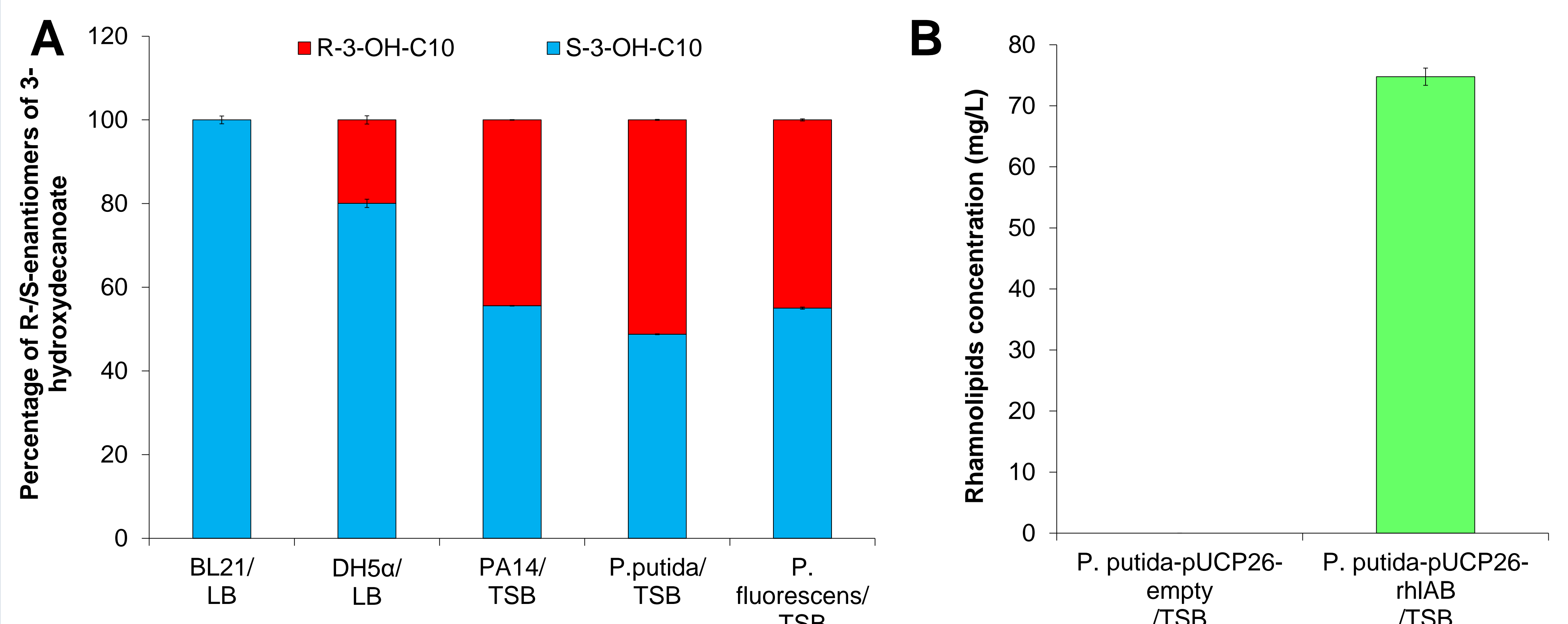
### *R*-specific ECH activity is in synchrony with RL production level by *P. aeruginosa* in different culture conditions



**Figure 1.** Rhamnolipids production by *P. aeruginosa* PA14 in optimized minimal salts medium (MSM) is about 20 times that in rich Tryptic Soy Broth (TSB) medium (Fig. 1A). Synchronously, the *in vitro* *R*-specific ECH activity (supplying *R*-3-hydroxyacyl-CoA precursors of RL) in cells of *P. aeruginosa* PA14 cultivated in MSM is two times higher than that when cultivated in rich medium (Fig. 1B)

## Results-2

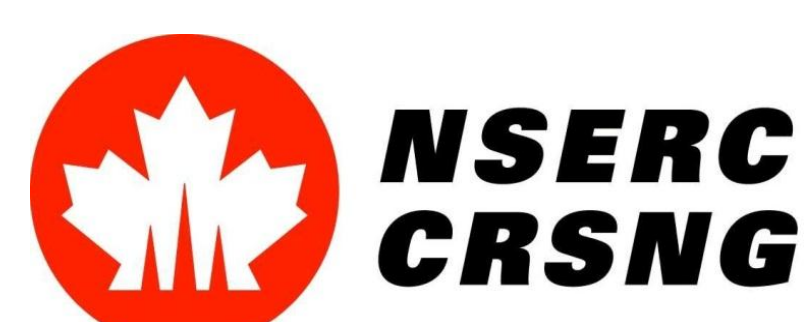
### Chiral lipid profiling as a tool for identification of candidate heterologous hosts for RL expression: e.g. *P. putida* KT2440



**Figure 2.** *In vitro* assay of *R*/*S*-ECH activities in total cell lysates of different strains shows variable degrees of *R*-specific ECH activities which is null in *E. coli* BL21 and detectable in *P. putida* (KT2440) (Fig. 2A). Accordingly, the latter strain is rather suggested to be supplying lipid precursors of RL and hence a probable candidate host for RL expression as proved experimentally in Fig. 2B.

## Acknowledgements

This study was supported by NSERC Discovery grant No. 312478 to E.D. A.M.A-M. is recipient of a Vanier - Canada Graduate Scholarship, and of a tuition fees grant from the Ministry of Higher Education and Research, Egypt. E.D. holds a Canada Research Chair.



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## Conclusions

- We demonstrate an efficient tool of chiral HPLC-tandem MS to estimate the net ECH activity in cell lysates and determine its type (*R*/*S*-specificity).
- For maximum and/or heterologous expression of rhamnolipids, it is a prerequisite that the host is naturally supplying the two precursors of Rhamnolipids (RL); L-rhamnose and *R*-3-hydroxyalkanoate.
- The provided tool helps in the identification of the physiological conditions with maximal supply of the latter precursor, *R*-3-hydroxyalkanoate, for maximum production of RL.
- Finally, the method is useful in the identification of potential candidate hosts for heterologous expression of RL based on the nature of their *R*-specific ECH activity.